Table I. Relative Quantum Yields for Photoisomerization and Cyclization^a

Compd	Reaction	Solvent	Wave- length, nm	Catalyst	Rel quan- tum ^b yield
3 ^c	Cyclization	$(C_{2}H_{5})_{2}O$	300	None	0.05
1 c	Trans → cis	$(C_2H_5)_2O$	300	None	0.90
1 c	Cyclization	$(C_2H_5)_2O$	300	None	0.02
1 c.d	Cyclization	$(C_{2}H_{5})_{2}O$	300	CuCl	0.20
1 c, d	Cyclization	CHCl ₃	350	Ph ₂ CO	0.10
5°	Trans → cis	$(C_{2}H_{5})_{2}O$	300	None	0.90

^a Standard: photoisomerization (trans \rightarrow cis) of 1,3-propanedioldicinnamate = 1.0 in diethyl ether at 300 nm. ^b Values are extrapolated to zero conversion. These values can be converted to absolute quantum yields by the use of 0.47 for the quantum yield of the standard.⁶ ^c Concentration 5×10^{-5} M. ^d Concentration 10^{-4} M.

which has the structure 5 was found not to undergo internal photocycloaddition under any of the experimental conditions that were tried. Reaction 1 is of interest from two points of view which follow.



(i) One concerns the factors which control the probability of a successful encounter between two cinnamate groups in a given molecule. The two important considerations according to the present investigation are the rate of a diffusive encounter between the chromophoric groups and the lifetime of the excited state of the molecule. Direct irradiation of 1 or 3 or 5 seems to lead to reaction from a singlet state as triplet sensitization gives quite different results. In this singlet state, trans \rightarrow cis isomerization proceeds with about equal efficiency in all three instances (which is reasonable), but cyclization is twice as efficient in 3 (s = 8) as in 1 (s = 17), while 5 which has nearly the same separation between the chromophores as 3 does not cyclize at all to any detectable extent. The decrease in reactivity in going from 3 to 1 parallels the reported³ decrease in the quantum yields for the closure of bis anthroates with separations of 7 and 14 bonds and is attributable to decreased probability of an encounter between the ends of the chain with increasing chain length. The sharp contrast in behavior between 3 and 5 which have nearly the same separation suggest yet another consideration. Molecular models show that a [2 + 2] internal adduct of 5 would not suffer from angle strain whatever the stereochemistry of the addition may be, but severe limitations on its conformational mobility are placed by its *trans*-1,3-cyclobutane geometry. Therefore, separations between chromophores are comparable only when the geometries of the molecules are strictly similar.

For a given reactant molecule, the number of encounters between the ends is undoubtedly increased by going from a singlet to a triplet excited state. The marked effectiveness of triplet sensitizers on the photoreaction of various cinnamate esters (including polyvinyl cinnamate) is well documented in the literature.^{13,14} The data in Table I also bear this out. Cuprous chloride may also function by promoting the intersystem crossover through a heavy atom effect.15

(ii) A second point of interest is the stereochemistry of the addition in reaction 1. In the solid state it has been shown⁴ that a [2 + 2] photocycloaddition between cinnamate groups will give exclusively α -truxillic or β -truxinic acids or their derivatives. The conditions which govern the formation of one or the other have been elegantly worked out. In contrast, in solution, as already mentioned, the δ -truxinic ester is a product of the photocyclization of 3 and is the only product from 4. The present work suggests that the latter mode persists with even longer molecular separation between cinnamate groups. It may be noted that all three dicarboxylic acids are derived from trans-cinnamic acids. The stereochemistry that prevails in the photo cross linking of polyvinyl cinnamate which is usually irradiated as an amorphous film is an interesting question. In our earlier work,¹⁶ we had looked for only α -truxillic and β truxinic acids among the hydrolysis products of the photolyzed materials. After establishing the stereochemistry in 2, the earlier data were reexamined to see if we could have overlooked the presence of δ -truxinic acid. It was confirmed that an amorphous film of polyvinyl cinnamate gives only α -truxillic acid as we determined before. This indicates that the behavior of the cinnamate groups in the film is similar to those in a crystal rather than a fluid solution. The structure of the film deserves further examination.

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Sirohydrochlorin. Prosthetic Group of a Sulfite Reductase Enzyme and Its Role in the Biosynthesis of Vitamin B₁₂

Sir:

Recent work in these laboratories^{1,2} and independently at Cambridge³ and Stuttgart⁴ has confirmed the role of uro'gen III (1) in the biosynthesis of vitamin B_{12} (2). It has also been shown⁵ that, during the bioconversion of both uro'gen III (1)and the "ring C heptacarboxylic acid" (4) to cobryinic acid (3), formaldehyde can be trapped from the δ -meso (C-20) carbon of 1 and 4. The relatively low but intact conversion of 4 to 3 (ca.



Figure 1. The ${}^{13}C$ FT spectrum of [4- ${}^{13}C$]Ala-labeled sirohydrochlorin octamethyl ester with proton noise decoupling (main spectrum) and with off-resonance CW decoupling (inset) recorded on a Varian FT-80 instrument.



an order of magnitude less than the incorporation of 1 into 3) suggests reassessment of the location of the heptaacid (4) on the metabolic grid of the porphyrinogen-corrin pathway. In this paper we describe the characterization of a new isolate from a modified version of the *P. shermanii* cell-free system⁶ which has considerable bearing on the porphyrinogen-corrin connection.

A novel heme-like prosthetic group in a rather widespread class of enzymes which catalyze the six-electron reduction of sulfite to sulfide in *E. coli* (E.C. 1.8.1.1) was first characterized in 1973⁷ and named siroheme. Removal of the iron from this species afforded an orange fluorescent compound—sirohydrochlorin. By modifying our cell-free technique it has been possible to isolate from *P. shermanii* almost 1 mg of an orange fluorescent substance.

Inspection of the UV, CD, mass and ¹H NMR spectral data for the methyl ester of the *P. shermanii* metabolite and of sirohydrochlorin ester from *E. coli* NADPH-sulfite reductase leaves no doubt that the substances, which also show complete correspondence in TLC R_f values, are identical in every respect. The molecular constitution was also confirmed by high resolution mass determination of the molecular ion at m/e 974 which revealed the composition of C₅₀H₆₂N₄O₁₆ (974.4160).

As pointed out by Siegel et $al.^{7,8}$ the UV spectrum of sirohydrochlorin is diagnostic of the isobacteriochlorin class requiring the two reduced rings to be adjacent (see structures **5-8**).

In the ¹H NMR spectrum recorded at 270 MHz the 3 H singlet resonances at δ 1.87 and 1.93 are consistent with the known chemical shifts of methyl groups on the reduced ring of a chlorin.⁹⁻¹¹ Similarly a 2 H multiplet at δ 4.1 also falls within the range of shifts expected for methine hydrogens in such systems.⁹⁻¹² Decoupling experiments showed that the methylene hydrogens of the acetate moieties adjacent to the



Figure 2. The meso carbon resonances of the proton noise-decoupled ¹³C FT spectrum of [5-¹³C]Ala-labeled sirohydrochlorin octamethyl ester in CDCl₃.



reduced ring (δ 2.72–2.75) were not coupled to the δ 4.1 multiplet (or any other resonance). Structures **5** and **8** are, therefore, eliminated as they would require coupling between these two resonances. Furthermore, incubation of 60 mg of [4⁻¹³C]aminolevulinic acid (90% enriched) with the *P. shermanii* cell-free homogenate yielded a purified sample of sirohydrochlorin (400 μ g) whose proton-decoupled and off-resonance CW decoupled spectra are in Figure 1. It is clear that only structures **6** and **7** are compatible with the observation of two doublets (C-3 and C-8) (see insert, Figure 1), since structures **5** and **8** would exhibit only one such enriched sp³ carbon bearing hydrogen. Finally, incubation with [5⁻¹³C]aminolevulinic acid enabled us to show that sirohydrochlorin has

structure 7. The expected labeling pattern is as shown in Figure 2. In the proton-decoupled spectrum, the C-15 resonance is a triplet (J = 72 Hz) due to 1,2 coupling with two adjacent enriched sites. (A lower intensity doublet is also present owing to those molecules having only one adjacent enriched site.) The C-5 and C-10 resonances both occur as doublet of doublets due to 1,2 and 1,4 couplings while C-20 shows only a 1,4 interaction. The four-meso-hydrogen ¹H NMR resonances occurred as doublets (J = 183 Hz) at δ 8.54, 7.46, 7.36, and 6.78. As discussed by Bonnett et al.,^{12a} the upfield meso-hydrogen resonance of an isobacteriochlorin may be assigned to that between the reduced (methylated) rings and the downfield resonance to that between the nonreduced rings. Thus the δ 8.54 hydrogen would be coupled to C-15 in structure 7 and to C-20 in structure 6. The former case was confirmed by a selective heteronuclear decoupling experiment. Additionally, the δ 6.78 meso hydrogen was shown to be coupled to the upfield meso carbon which is therefore C-5.

The structural proposal (7) for sirohydrochlorin was examined by two sets of biosynthetic experiments. Specimens of sirohydrochlorin labeled (in $\sim 10\%$ radiochemical yield) by separate incubations⁶ with δ -[4-¹⁴C]aminolevulinic acid and S-[³H₃C]adenosylmethionine were purified chromatographically as the octamethyl esters, hydrolyzed, and reincubated as singly and doubly labeled species after sodium amalgam reduction $(7 \rightarrow 9)$. The singly labeled species was incorporated by the "corrin synthetase" 6 preparation into cobyrinic acid 3 (isolated as cobester) in 0.4-1.9% radiochemical yield. Intact incorporation of the reduced version (9) is clearly revealed by retention of the ${}^{3}H/{}^{14}C$ ratio (3.45) in the isolated cobester (3.41). When unreduced 7 was incubated with the corrin synthetase preparation, similar incorporations (0.3-2.8%) were observed. The absolute stereochemistry of 7 is therefore established via bioconversion to corrin.13

Other workers^{14,15} have recently isolated a metabolite from P. shermanii with similar UV and mass spectral characteristics to those of sirohydrochlorin. The Cambridge group postulated structure 7 for their metabolite after assuming that it was on the B_{12} pathway and further that ring C was not methylated. This paper¹⁶ shows unequivocally that sirohydrochlorin is indeed identical with a P. shermanii metabolite, that it has structure 7, and that it is an intermediate on the corrin pathway. The possibility that siroheme represents a prebiotic sulfate-reducing agent⁸ and, further, that both sirohydrochlorin and vitamin B_{12} producing anaerobic organisms predate the evolution of heme-synthesizing aerobes¹⁷ suggest that the reductive methylation of reduced porphyrins may be a phenomenon of considerable antiquity (three billion years).

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Enhanced Chemiluminescence from the Silica Gel Catalyzed Decomposition of a 1,2-Dioxetane

Sir:

In 1969 Kopecky and Mumford first demonstrated that the thermolysis of 1,2-dioxetanes yields electronically excited carbonyl products.¹ It has subsequently been shown that simple isolable 1,2-dioxetanes afford predominantly triplet excited states upon spontaneous decomposition in solution.² Transition metals,³ electron-rich olefins,⁴ and amines⁴ catalyze the cleavage of 1,2-dioxetanes. However, these processes give ground-state products and therefore result in reduced light emission. We now report the first observation⁵ of enhanced chemiluminescence associated with the catalytic decomposition of a stable, crystalline 1,2-dioxetane.

2,3-Di(2-anthryl)-1,4-dioxene (1,6 100 mg) was photooxygenated for 30 min at -78 °C in 100 mL of CH₂Cl₂ using 1 g of polymer-bound Rose Bengal⁷ as sensitizer with two 500-W tungsten-halogen lamps and a UV-cutoff filter. The reaction mixture was filtered and concentrated at 10 °C on a rotary evaporator to 50 mL. Addition of 1 mL of pentane and cooling to -25 °C gave pure 2 as a pale yellow solid in 51% yield: UV (o-xylene) λ_{max} 327 nm (log ϵ 3.68), 343 (3.83), 361 (3.93), 380 (3.82);^{8 1}H NMR (100 MHz, CDCl₃, 0 °C) δ 4.51 (m), 4.99 (m), 6.5-8.6 (m, aromatic). Additional 2 could be obtained by concentrating the reaction solution further;⁹ however, this material also contained cleavage product 3.

Thermolysis of the dianthryl-substituted 1,2-dioxetane 2 in o-xylene results in quantitative formation of the diester 3^{10} and is accompanied by light emission. The rates of decomposition of 2, the formation of 3, and the emission of light are first order and identical ($k = 6.56 \times 10^{-3} \text{ s}^{-1}$ at 84.1 °C). The